

Sub D > PEPTIDE ANTAGONISTS OF CGRP-RECEPTOR SUPERFAMILY AND
METHODS OF USE

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Field of the Invention

5 This invention relates to the field of vasoactive compounds and their antagonists. In particular, this invention relates to antagonists of the vasoactive peptide CGRP and other members of the CGRP superfamily.

Background of the Invention

10 This invention supported in part by NIH Grant No. HL51131. The government may have certain rights to this invention.

INS A2 >

15 The calcitonin gene related peptide (CGRP) is a sensory neuropeptide with potent vasodilatory and cardiotonic action as described in U.S. Pat. No. 4,530,838 to Evans et al. The peptide exists in two forms (denoted α and β). α -CGRP is produced by the calcitonin gene (Amara et al. *Nature* 298:240-244, 1982 and Rosenfeld et al. *Nature* 304:129-135, 1983) while β -CGRP is the product of a separate gene (Amara et al. *Nature* 298:240-244, 1985 and Steenbergh et al. *FEBS Lett.* 183:403-407, 1985). The human β -form and α -form differ by three amino acids.

20 CGRP is concentrated in those areas of the body receiving sensory input from the dorsal horn with limited amounts associated with autonomic input. The peptide is present in the brain in the nuclei of sensory and motor cranial nerves and in cell bodies in the hypothalamus, preoptic area, ventromedial thalamus, hippocampus, and the like. CGRP is found in both sensory and motor nerves of the peripheral nervous system. The peptide is found in the skin, blood vessels, heart, 25 gastrointestinal tract, tongue, esophagus, pancreas, salivary glands, lungs, kidney and other organs (Poyner, D. *Pharmac. Ther.* 56:23-51, 1992).

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The release of CGRP from sensory nerve endings in inflammatory reactions can result in the local acceleration of microhemodynamic changes including vasodilation and permeability of the microcirculation resulting in plasma

exudation and the release of humoral factors and inflammatory cells to the site of injury. CGRP has been used as a vasodilator in animal models of subarachnoid hemorrhage and in trials involving human subjects with congestive heart failure. CGRP administration produced hypotension associated with moderate tachycardia in
5 hypertensive humans (Jian et al. *Chin. Med. J.* 102:897-901, 1989). CGRP has also been used as a potent dilator of the coronary circulation (Ezra et al., *Eur. J. Pharmacol.*, 1987). In contrast to nitrates, which have also been used as vasodilators, CGRP results in dilation by both endothelium-dependent and endothelium-independent mechanisms. Also, in contrast to nitrates, such as sodium
10 nitroprusside, tolerance to CGRP has not been noted (Foulkes et al. *Regul. Pept.* 25:25-36, 1989). CGRP has been demonstrated to improve the ability of patients to participate in exercise programs in patients with chronic stable angina (Uren et al. *Cardiovasc. Res.* 27:1477-1481, 1993).

CGRP has a number of problems as a therapeutic. CGRP is
15 nonselective, inactive in oral form, generally has a short duration of action and has a number of side effects that can include uncontrolled hypotension (Feuerstein et al. *Can. J. Physiol. Pharmacol.* 73:1070-1074, 1995).

CGRP has been implicated in migraines, diabetes, sepsis and inflammation. Migraines are noted for the strength of the headache that ensues with
20 its pathology. Most believe that the headache associated with migraines results from the profound cerebral vasodilation. CGRP containing nerve fibers innervate cerebral and dural vessels where CGRP is believed to prolong vasodilation. (Moskowitz *Trends Pharmacol. Sci.* 13:307-311, 1992). Elevated CGRP was found in the jugular vein blood of patients with migraines during a period where the
25 patients complained of migraine symptoms, including headaches. For these reasons, CGRP antagonists have been proposed as a method for blocking cerebrovascular CGRP receptors and thus blocking the vasodilation causing migraine.

CGRP has also been postulated to be a potent indirect antagonist of insulin effects on glucose metabolism and CGRP was shown to produce insulin resistance in rat studies (Molina et al. *Diabetes* 39:260-265, 1990). For this reason CGRP has recently been implicated in Type II diabetes mellitus and to abnormalities associated with carbohydrate metabolism and hyperglycemia. CGRP has also been implicated in the hemodynamic derangements associated with endotoxemia and sepsis resulting from a variety of infectious diseases. Animals exposed to lipopolysaccharide (LPS) had elevated levels of CGRP and this coincided with hypotension and tachycardia in these animals (Joyce et al. *Surgery* 108:1097-1101, 1990 and Griffin et al. *Circ. Shock* 38:50-54, 1992).

CGRP binds to a number of different receptors, some of which have been characterized. Radioligand binding studies to assess CGRP affinity for CGRP receptors is well known in the literature (Poyner, D.R. *Pharmac. Ther.* 56:23-51, 1992). As stated in Poyner et al., a problem associated with studies to identify CGRP receptors is that lack of suitable CGRP receptor binding analogs and it is accepted that the use of CGRP antagonists is a useful way of classifying CGRP receptors. The art recognizes that there are a limited number of antagonists and that it would be desirable to have more CGRP antagonists to further classify and understand CGRP activity.

Molecules that compete for the CGRP receptor are known. These include, for example, [Tyr⁰]CGRP(28-37) and CGRP(8-37). Other molecules that compete for the CGRP receptor include peptides comprising the sequence of CGRP but that lack at least the first five amino acids of the CGRP amino acid sequence. [Tyr⁰]CGRP(28-37) was able to antagonize all forms of CGRP tested but with different potencies. Other molecules that compete for the CGRP receptor are provided elsewhere in this disclosure.

~~β-CGRP antagonists includes peptides from CGRP including amino acids 8-37 of β-CGRP (Chiba et al. *Am. J. Physiol.* 1989) having the amino acid sequence: THRLAGLLSRSGGMVKSNFVPTNVGSKAF (SEQ ID NO:1) and~~

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peptides from α -CGRP including amino acids 8-37 and having the amino acid sequence THRLAGLLSRSGGMVKS NFVPTNYGSKAF. β -CGRP(8-37) (SEQ ID NO:2) has been used to counteract the effects of CGRP. For example, CGRP(8-37) has been shown to reverse the hypotension and tachycardia produced by administration of LPS to rats (Huttemeir, et al. *Am. J. Physiol.* 265:H767-H769, 1993). In addition, CGRP(8-37) has some activity against amylin (Gardiner et al. *Diabetes* 40:948-951, 1991). The affinity for CGRP(8-37) varies between tissues. For example, data indicates that the affinity of CGRP(8-37) for mesenteric artery, kidney, heart and skeletal muscle is somewhat higher than the affinity of CGRP(8-37) for adipocytes and descending colon (Poyner, D. *Trends in Pharm. Sci.* 16:424-428, 1995).

Brief Description of the Figures

Figure 1A is a graph demonstrating the inhibition of α -CGRP binding to coronary artery membranes in the presence of exemplary peptides, according to this invention. The symbol ■ indicates h- α CGRP (8-37) Δ N-benzyl-h- α -CGRP (8-37) \diamond N-benzoyl-h- α -CGRP (8-37) \circ dibenzyl-h- α -CGRP (8-37). Figure 1B is a graph showing the inhibition of CGRP binding by the β CGRP (8-37) derivatives to coronary artery membranes in the presence of exemplary peptides according to this invention. The symbol ■ indicates h- β CGRP (8-37) Δ N-benzyl-h- β CGRP (8-37) \diamond N-benzoyl-h- β CGRP (8-37) \circ dibenzyl-h- β CGRP (8-37).

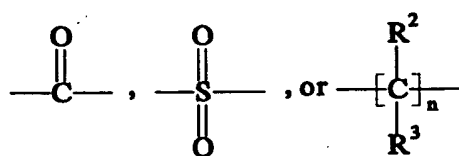
Figure 2 illustrates the antagonist effect of α CGRP(8-37) modified peptides on h- α CGRP-induced relaxation of capsaicin-treated pig coronary artery. Figure 2A illustrates the antagonistic effect of h- α CGRP(8-37). The symbol \square indicates Control + h- α CGRP (8-37) ■ 3×10^{-6} M \blacktriangle 1×10^{-5} M \bullet 3×10^{-5} M. Figure 2B illustrates the antagonistic effect of N-benzoyl-h- α CGRP(8-37). The symbol ■ indicates Control + N-benzoyl-h- α CGRP (8-37) \square 1×10^{-7} M Δ 3×10^{-7} M \diamond 1×10^{-6} M \circ 3×10^{-6} M. Figure 2C illustrates the antagonistic effect of N-benzyl-h- α CGRP(8-37). The symbol ■ indicates Control + N-benzyl-h- α CGRP (8-37) Δ 1

$\times 10^{-6} \text{ M}$ $\diamond 3 \times 10^{-6} \text{ M}$ $\circ 1 \times 10^{-5} \text{ M}$. Figure 2D illustrates the antagonistic effect of dibenzyl-h- α CGRP(8-37). The symbol \blacksquare indicates Control + dibenzyl-h- α CGRP (8-37) $\Delta 3 \times 10^{-7} \text{ M}$ $\diamond 1 \times 10^{-6} \text{ M}$ $\circ 3 \times 10^{-6} \text{ M}$.

Summary of the Invention

- 5 The present invention relates to vasoactive peptides having the general formula: $\text{R}^1\text{-X-Z}$

wherein Z is a vasoactive peptide, R¹ is an organic group, X is



, and wherein R² and R³ are

independently H or an organic group and n is a whole integer between 1 and 10.

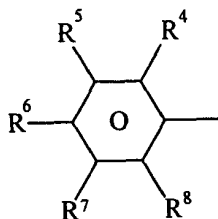
- 10 In a preferred embodiment, Z is a peptide fragment of CGRP. In one embodiment the amino acid sequence of CGRP is SEQ ID NO:3 and in another the amino acid sequence of CGRP is SEQ ID NO:4. Preferably the CGRP is human CGRP and preferably either α -CGRP or β -CGRP. In a preferred embodiment the peptide fragment is CGRP(8-37), Tyr^o CGRP(28-37), or CGRP(12-37). In another
- 15 embodiment the peptide fragment is a CGRP antagonist and preferably selected from the group of peptides or peptide fragments from amylin or adrenomedullin that bind to one or more CGRP receptors. In another embodiment, the peptide fragments are peptide fragments with CGRP antagonist activity selected from CGRP receptor
- 20 binding peptides preferably having at least 15 amino acids from the amino acid sequences of SEQ ID NOS:5-19.

- Also preferably, R¹ is an aromatic group, a heterocyclic group or an alkyl group and R² and R³ are independently H, an aromatic group or an alkyl group. In one embodiment, R¹ is a C1-C18 aromatic group, or a C1-C4 alkyl group and in another embodiment, R¹ is a fluoroalkyl group. In another embodiment, R¹ is a C5-
- 25 C10 aromatic group, a C5-C9 heterocyclic group or a C1-C18 alkyl group.

Preferably R^2 and R^3 are independently H, a C1-C4 alkyl group or a phenyl moiety. Preferably, R^2 and R^3 are independently H or a C5-C10 aromatic group or a C1-C4 alkyl group.

In one embodiment, R^1 has the general formula:

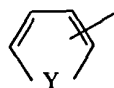
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and R^4 - R^8 are each independently selected from the group of H, fluoro, chloro, bromo, iodo, nitro, nitrile (cyano), amino, N-methyl amino, N,N-dimethyl amino, hydroxy, methoxy, thiomethoxy (S-methyl), methyl, ethyl, n-propyl, iso-propyl, -butyl, iso-butyl, sec-butyl, tert-butyl, trifluoromethyl, trifluoromethoxy, vinyl, acetamido, phenyl, tolyl, and methoxyphenyl. In a preferred embodiment, R^6 is trifluoromethyl and one or more of R^4 , R^5 , R^7 and R^8 is F.

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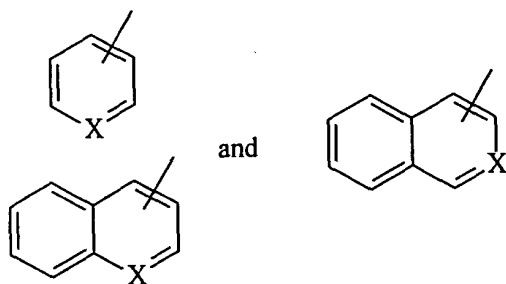
In another embodiment of the invention, R^1 is



and Y is selected from the group consisting of O, NH, and S.

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In yet another embodiment, R^1 is selected from the group consisting of:



and X is C or N.

In another aspect of this invention, the peptides of this invention are selected from the group consisting of N- α -benzoyl- α -CGRP receptor antagonist peptides, N- α -benzyl- β -CGRP receptor antagonist peptides, N- α -benzoyl- β -CGRP
5 receptor antagonist peptides, N- α -benzyl- α CGRP receptor antagonist peptides, N- α -benzyl-[(4'-benzyl-His¹⁰)- α -CGRP receptor antagonist peptides, [(4'-benzyl-His¹⁰)- β -CGRP receptor antagonist peptides, N, N-dibenzyl- α -CGRP and N,N-dibenzyl- β -CGRP.

The invention also relates to a method for inhibiting CGRP binding to
10 one or more CGRP receptors comprising the step of contacting an effect amount of a peptide of this invention with the CGRP receptor wherein R¹ of the peptide is an aromatic group, a heterocyclic group or an alkyl group and R² and R³ of the peptide are independently H, an aromatic group or an alkyl group. The receptor can be cell free or cell associated. The receptor can be in a cell in culture or in a cell as part of a
15 tissue of an animal, including humans.

The invention further relates to an assay for identifying CGRP antagonists comprising the step of: combining at least one peptide of this invention and a candidate CGRP antagonist with a CGRP receptor and comparing binding of the peptide to the CGRP receptor and binding of the candidate antagonist to the
20 CGRP receptor wherein binding of the candidate antagonist to the CGRP receptor in the presence of the peptide identifies a CGRP antagonist.

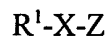
The invention also relates to a method for inhibiting CGRP activity comprising the step of administering an effective amount of a peptide of this invention to a cell wherein the cell comprises a CGRP receptor.

25 In yet another aspect of this invention, the invention relates to a method for identifying a CGRP receptor in a cell sample comprising the step of detecting binding of a peptide of this invention to a cell and isolating and/or characterizing the receptor.

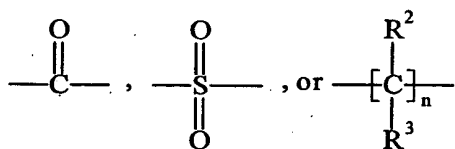
Detailed Description of the Preferred Embodiments

The present invention relates to modified peptides that serve as vasoactive peptide antagonists for the CGRP receptor. In a preferred embodiment, this invention relates to peptides with amino terminal modifications wherein the peptide functions as a CGRP antagonist.

In one embodiment, this invention relates to vasoactive peptides with the following general formula:



wherein Z is a vasoactive peptide fragment, R^1 is an organic group, X is



and wherein R^2 and R^3 are independently H or an organic group and n is a whole integer between 1 and 10.

The term "vasoactive peptide" refers to peptides that are capable of causing vasoconstriction or vasodilatation of blood vessels and a peptide capable of binding to a CGRP receptor refers to peptides, preferably of at least 15 amino acids in length that have CGRP receptor binding activity. For purposes of this invention, a CGRP receptor is an isolated or cell associated receptor with CGRP binding activity.

A variety of vasoactive peptides or peptide fragments that function as CGRP antagonists are known in the art and these include, but are not limited to, CGRP receptor- binding peptide fragments of CGRP, including the α and β forms of CGRP and peptides of adrenomedullin and amylin. CGRP has been isolated from a variety of animals including, but not limited to humans (β -CGRP, SEQ ID NO:3; α -CGRP; SEQ ID NO:4, Poyner, D.R. *Pharmac. Ther.* 56:23-51, 1992), rats (β -CGRP, SEQ ID NO:5; α -CGRP. SEQ ID NO:6, Poyner, *supra*), chickens (SEQ ID NO:7, Poyner, *supra*), rabbits (SEQ ID NO:8, Eysselein et al. *Peptides* 12:289-295, 1991),

pigs (SEQ ID NO:9, Kimura, S. et al. *Neuropeptides* 9:75-82, 1987), sheep (SEQ ID NO:10, Miyata et al. *Biochem. Biophys. Res. Commun.* 187:1474-1479, 1992), cows (SEQ ID NO:11 Collyear, K. et al. *J. Mol. Endocrinol.* 6:147-152, 1991), salmon (SEQ ID NO:12, Jansz, et al. *Ann. N. Y. Acad. Sci.* 657:63-69, 1992) and frogs (SEQ ID NO:13 Esneu et al. *Endocrinol.* 135:432-430, 1994). Adrenomedullin has been isolated from a variety of sources including human (SEQ ID NO:14 Kitamura, K. et al. *Biochem. Biophys. Res. Commun.* 195:921-927, 1993) and rat (SEQ ID NO:15 Sakata, J. et al. *Biochem. Biophys. Res. Commun.* 195:921-927, 1993). Amylin has also been isolated from a variety of sources including, but not limited to, human (SEQ ID NO:16 Westermark, P. et al. *Proc. Natl. Acad. Sci. USA* 84:3881-3885, 1987) and rat (SEQ ID NO:17 Leffert, J.D. et al. *Proc. Natl. Acad. Sci. USA* 86:3127-3130, 1989).

Antagonists of the CGRP receptor include a variety of peptides including peptide fragments from CGRP peptides including, but not limited to , CGRP (8-37), CGRP (28-37) including Tyr^oCGRP (28-37), and CGRP (12-37). Other CGRP antagonists include h- α -CGRP (9-37), h- α -CGRP (10-37), h- α -CGRP (11-37) (Mimeault, M. et al., *J. Med. Chem.* 35:2163-2168, 1992). Still other CGRP antagonists include [Ala⁹]-h- α -CGRP (8-37), [Ala¹⁰]-h- α -CGRP (8-37), [Ala¹¹]-h- α -CGRP (8-37), and [Ala¹²]-h- α -CGRP (8-37). Additional CGRP antagonists include h- α -CGRP(19-37), h- α -CGRP(23-37) and acetyl-h- α -CGRP(19-37) (Rovero, P. et al. *Peptides* 13:1025-1027, 1992).

Amylin antagonist peptides are known and a number of these with CGRP receptor binding activity are provided in U.S. Pat. No. 5,625,032 to Gaeta et al., and U.S. Pat. No. 5, 580,953 to Albrecht et al. Preferred amylin antagonist peptides include:

human amylin (8-37)

HATQRLANFLVHSSNNFGAILSSTNVGSNTY-NH₂ (SEQ ID NO:18),

rat-amylin(8-37)

H-ATQRLANFLVRSSNNLGPVLPPTNVGSNTY-NH₂) SEQ ID NO:19),
and Acetyl-Rat-Amylin (8-37) (Deems et al. *Biochem. Biophys. Res. Commun.*
181:116-120, 1991).

Still other amylin antagonists that can be tested for CGRP receptor
5 binding activity include:
AC187 (SEQ ID NO:20)
(Acetyl-VLGKLSQELHKLQTYPRTNTGSNTY-NH₂, Beaumont et al. *Br. J.*
Pharmacol. 115:713-715, 1995),
AC253 (SEQ ID NO:21)
10 (Acetyl-LGRLSQELHRLQTYPRTNTGSNTY-NH₂), and
AC625 (SEQ ID NO:22)
(Acetyl-ATQRLANELVRLQTYPRTNVGSNTY-NH₂ both Prickett, K.S. et
al. in *Peptides: Chemistry and Biology*, eds. Kaumaya, P.T.P and Hodges, S. .
Mayflower Scientific Ltd., Kingswinford, UK. 1996).

15 Preferred adrenomedullin-derived antagonists include:
h-adrenomedullin (22-52) (SEQ ID NO:23)
(TVQKLAHQIYQFTDKDKDNVAPRSKISPQGY-NH₂, Watanabe, et al.
Endocrinol. 135:2454-2458, 1994 and Champion et al. *Am. J. Physiol.* 272:R234-
242, 1997),

20 In addition, the modifications of this invention can be incorporated
into other polypeptides with vasoactivity and having CGRP receptor binding activity,
such as relaxin, a molecule structurally related to amylin (Cooper et al., *Proc. Natl.*
Acad. Sci. USA 85:7763-7766, 1988). Moreover, substituted peptides of amylin,
CGRP or other vasoactive peptides are described in U.S. Application Serial No.
25 275,475.

In general, the vasoactive peptides of this invention include a
carboxamide at the C-terminus. Alternatively, the peptides of this invention can
include a free carboxyl group at the terminus. Abbreviations for peptide termini are
as follows: "H-" refers to a free amino group, "-OH" refers to a free carboxyl group

and "-NH₂" refers to a carboxamide. The term "vasoactive peptide" as used herein refers to peptides with physiological activity, particularly, but not necessarily solely, directed in activity to the vasculature system and preferably peptides with CGRP antagonist activity. In general, the peptides of this invention exhibit greater activity
5 when a carboxamide is positioned at the terminus of the peptide. Methods for preparing peptides with C-terminal amide groups are known in the art and, in one example, described in U.S. Pat. No. 5,503,989 to Bibbs et al.

The amino acid designations used throughout this patent application include the standard amino acid designations: A or Ala for Alanine, C or Cys for
10 Cysteine, D or Asp for Aspartic acid, E or Glu for Glutamic acid, F or Phe for Phenylalanine, G or Gly for Glycine, H or His for Histidine, I or Ile for Isoleucine, K or Lys for Lysine, L or Leu for Leucine, M or Met for Methionine, N or Asn for Asparagine, P or Pro for Proline, Q or Gln for Glutamine, R or Arg for Arginine, S or Ser for Serine, T or Thr for Threonine, V or Val for Valine, W or Trp for
15 Tryptophan and Y or Tyr for Tyrosine.

R¹ in the formula R¹-X-Z is preferably an organic group. In a preferred embodiment, R¹ is an aromatic group, a heterocyclic group or an alkyl group. In a particularly preferred embodiment, R¹ is a C₅-C₁₀, a C₅-C₁₀ heterocyclic group and more preferably a C₅-C₉ heterocyclic group, or a C₁-C₁₈
20 alkyl group and more preferably a C₁-C₄ alkyl group. In one embodiment, R¹ is a fluoroalkyl.

As used herein, the term "organic group" refers to a hydrocarbon group that is classified as an aliphatic group an aromatic group, a cyclic group, or a combination of aliphatic and cyclic groups (e.g., alkaryl and aralkyl groups). In the
25 context of the present invention, the term "aliphatic group" refers to a saturated or unsaturated linear or branched hydrocarbon group. This term is used to encompass alkyl, alkenyl, and alkynyl groups, for example. The term "alkyl group" means a saturated linear or branched hydrocarbon group including, for example, methyl, ethyl, isopropyl, t-butyl, heptyl, dodecyl, octadecyl, amyl, 2-ethylhexyl, and the like.

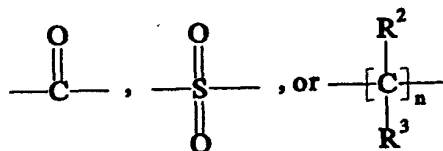
The term "alkenyl group" refers to an unsaturated, linear or branched hydrocarbon group with one or more carbon-carbon double bonds, such as a vinyl group. The term "alkynyl group" refers to an unsaturated, linear or branched hydrocarbon group with one or more carbon-carbon triple bonds. The term "cyclic group" refers to a closed ring hydrocarbon group that is classified as an alicyclic group, aromatic group, or heterocyclic group. The term "alicyclic group" means a cyclic hydrocarbon group having properties resembling those of aliphatic groups. The term "aromatic group" or "aryl group" refers to mono- or polynuclear aromatic hydrocarbon group. The term "heterocyclic group" refers to a closed ring hydrocarbon in which one or more of the atoms in the ring is an element other than carbon (e.g., nitrogen, oxygen, sulfur, etc.).

As is well understood in this technical area, a large degree of substitution is not only tolerated, but is often advisable. Substitution is anticipated on the compounds of the present invention. As a means of simplifying the discussion and recitation of certain terminology used throughout this application, the terms "group" and "moiety" are used to differentiate between chemical species that allow for substitution or that may be substituted and those that do not allow or may not be so substituted. Thus, when the term "group" is used to describe a chemical substituent, the described chemical material includes the unsubstituted group and that group with O, N, or S atoms, for example, in the chain as well as carbonyl groups or other conventional substitution. Where the term "moiety" is used to describe a chemical compound or substituent, only an unsubstituted chemical material is intended to be included. For example, the phrase "alkyl group" is intended to include not only pure open chain saturated hydrocarbon alkyl substituents, such as methyl, ethyl, propyl, t-butyl, and the like, but also alkyl substituents bearing further substituents known in the art, such as hydroxy, alkoxy, alkylsulfonyl, halogen atoms, cyano, nitro, amino, carboxy, etc.. Thus, "alkyl group" includes ether groups, haloalkyls, fluoroalkyls, nitroalkyls, carboxyalkyls, hydroxyalkyls, sulfalkyls, etc. On the other hand, the phrase "alkyl moiety" is

limited to the inclusion or only pure open chain saturated hydrocarbon alkyl substituents, such as methyl, ethyl, propyl, t-butyl, and the like.

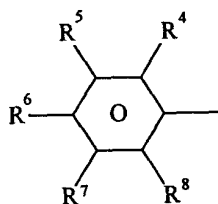
In the vasoactive peptides of this invention, having the general formula R^1-X-Z , X is preferably

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wherein R^2 and R^3 are independently H or an organic group and n is a whole integer from about 1 to about 20, preferably a whole integer from about 5 to about 20 and more preferably a whole integer from about 1 to about 4. In a preferred embodiment, R^2 and R^3 are independently H, an aromatic group, or an alkyl group and in a particularly preferred embodiment, R^2 and R^3 are independently H or a C5-C10 aromatic group or a C1-C18 alkyl group and more preferably a C1-C4 alkyl group. In yet another preferred embodiment, R^2 and R^3 are independently H, a lower alkyl moiety (e.g., about a C1-C4 alkyl) or a phenyl moiety.

10 In a preferred embodiment of this invention R^1 is an aromatic group, a heterocyclic group or an alkyl group and R^2 and R^3 are independently H, an aromatic group or an alkyl group. In one aspect of this embodiment, R^1 has the general formula:



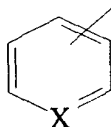
15 In a preferred embodiment of this invention R^4-R^8 are each independently selected from the group of H, fluoro, chloro, bromo, iodo, nitro, nitrile

(cyano), amino, N-methyl amino, N,N-dimethyl amino, hydroxy, methoxy, thiomethoxy (S-methyl), methyl, ethyl, n-propyl, iso-propyl, -butyl, iso-butyl, sec-butyl, tert-butyl, trifluoromethyl, trifluoromethoxy, vinyl, acetamido, benzyl, toluyl, and methoxybenzyl. In one preferred embodiment, R⁶ is trifluoromethyl and one or more of R⁴, R⁵, R⁷ and R⁸ are F.

In a further embodiment, R¹ is monocyclic, including, for example, both five-membered rings and six-member rings. A preferred five member ring with the general formula:



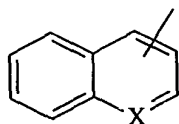
preferably includes Y as O, NH or S. The diagonal line extending from the center of five-membered ring and from the center of the rings of the structures depicted below indicates that the substituent -X-Z can be covalently attached to the ring at any of the carbon atoms that form the ring. In an embodiment where R¹ is a six-membered ring, preferably R¹ has the formula:



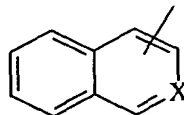
and preferably X is CH or N.

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In another embodiment of this invention the invention R¹ is a bicyclic ring having one of the following formulas:



and



wherein X is either N or C.

Preferred peptide modifications of this invention include, but are not limited to N- α -benzyl-Z, and N- α -benzoyl-Z, wherein Z is a peptide fragment capable of binding to a CGRP receptor. Preferred benzyl analogues include, but are not limited to, N-2-furanyl-Z, N-3-furanyl-Z, N-2-pyrrolyl-Z, N-3-pyrrolyl-Z, N-2-thiophenyl-Z, N-3-thiophenyl-Z, N-2-pyridyl-Z, N-3-pyridyl-Z, N-4-pyridyl-Z, N-1-naphthyl-Z, N-2-naphthyl-Z, N-2-quinolinyl-Z, N-4-quinolinyl-Z, N-8-quinolinyl-Z, N-1-isoquinolinyl-Z, N-3-isoquinolinyl-Z, R-N- α -methylbenzyl-Z, S-N- α -methylbenzyl-Z, α,α -dimethylbenzyl-Z, N-diphenylmethyl-Z, N-trityl-Z, and [D-Phe⁰]-Z. Other preferred benzoyl analogues of the peptide fragments of this invention include, but are not limited to N-2-furanoyl-Z, N-3-furanoyl-Z, N-2-pyrroloyl-Z, N-3-pyrroloyl-Z, N-2-thiophenoyl-Z, N-3-thiophenoyl-Z, N-2-thiophenoyl-Z, N-3-thiophenoyl-Z, N-2-pyridoyl-Z, N-3-pyridoyl-Z, N-4-pyridoyl-Z, N-1-naphthoyl-Z, N-2-naphthoyl-Z, N-2-quinolinoyl-Z, N-3-quinolinoyl-Z, N-4-quinolinoyl-Z, N-8-quinolinoyl-Z, N-1-isoquinolinoyl-Z, and N-3-isoquinolinoyl-Z. Still other preferred peptide modifications of this invention include N-methanesulphonyl-Z, N-trifluoromethanesulphonyl-Z, N-benzenesulphonyl-Z, N-toluenesulphonyl-Z, N-4-methoxybenzenesulphonyl-Z, N-mesitylenesulphonyl-Z, N-4-trifluorobenzenesulphonyl-Z and N-4-trifluoromethoxybenzenesulphonyl-Z.

Preferred vasoactive peptides fragments are peptide fragments of CGRP with CGRP antagonist activity, that is the peptide fragments of this invention inhibit CGRP activity. Preferably the peptides inhibit CGRP activity by at least 25% and preferably inhibit CGRP activity by at least 50%. Particularly preferred peptide fragments of this invention are peptide fragments of α -CGRP or β -CGRP, and preferably human α -CGRP or β -CGRP. Preferred peptide fragments of CGRP include CGRP (8-37).

The peptides of this invention can be prepared using methods known in the art. Exemplary methods for preparing the peptides of this invention are provided in Example 1. For example, a number of peptides according to this invention can be assembled on MBHA resin using the methodology of Smith, D.D.

et al. *J. Med. Chem.* 36:2536-2541, 1993. Those of ordinary skill in the art of peptide and protein modification can prepare the other peptide modifications of this invention without undue experimentation.

In one embodiment of this invention, the invention relates to a
5 method for inhibiting CGRP binding to one or more CGRP receptors by contacting an effective amount of a peptide of this invention with the CGRP receptor. This method can be used *in vitro*, for example in assays to identify and/or isolate CGRP receptors or with intact cells either *in vitro* or *in vivo* to inhibit the effects of CGRP binding to its receptor. As an example of an assay to determine the ability of the
10 peptides of this invention to compete with CGRP receptors, Example 2 illustrates an assay to determine whether a particular peptide modification of this invention can inhibit CGRP binding to a CGRP receptor.

Binding assays used to identify whether or not a particular peptide would inhibit CGRP binding to its receptor have been carried out using rat brain
15 (Dennis, et al., *J. Pharmacol. Exp. Ther.* 254:123-128, 1990, van Rossum, et al. *J. Pharmacol. Exp. Ther.* 269:846-853, 1994), spleen (Dennis, et al. *J. Pharmacol. Exp. Ther.* 254:123-128, 1990) and vas deferens (Mimeault, et al. *J. Pharmacol. Exp. Ther.* 258:1084-1090, 1991), guinea-pig atrium and vas deferens brain (Dennis, et al. *J. Pharmacol. Exp. Ther.* 254:123-128, 1990, van Rossum, et al. *J. Pharmacol. Exp. Ther.* 269:846-853, 1994), human neuroblastoma cells SK-N-MC (Rist et al. *J. Med. Chem.* 41:117-123, 1998) and pig brain, lung (Aiyar et al. *J. Neurochem.* 65:1131-1138, 1995) and kidney (Aiyar et al. *Endocrinology* 129:965-969, 1991).
20

Alternatively, the modified peptides of this invention, including those disclosed in the examples, can be used to identify other CGRP receptors or
25 alternatively, the assays of the present invention can be used to test and compare the efficacy of other CGRP antagonists. For example, the K_i for a specific peptide in binding to a specific type of CGRP receptor is a constant. If the K_i value for the same peptide is different in one tissue compared to another tissue then this is evidence for two different receptors in these tissues.

Where the peptides of this invention are used to identify other CGRP antagonists, the peptides of this invention can be used in competition assays with candidate antagonists for example, using either labeled peptide or labeled candidate antagonist, to assess preferential binding of the receptor to a peptide of this invention or to a test antagonist. The peptides of this invention or labeled candidate antagonist can be radiolabeled, labeled with a fluorescent tag, biotinylated or otherwise tagged and/or labeled using methods known in the art.

CGRP has been implicated in a variety of diseases and pathologies as has been described in the background section of this disclosure. CGRP acts as an antagonist of insulin action and CGRP is a potent vasodilator. Activity of CGRP is mediated through binding of CGRP to one or more CGRP receptors. For purposes of this disclosure, the term CGRP-receptor superfamily refers to the class of cell receptors that bind CGRP. The CGRP antagonist, CGRP(8-37) is a known antagonist but does not appear to consistently bind strongly to one or more CGRP receptors. Exemplary peptides of this invention have demonstrated an increase in binding affinity of about 65 fold over that reported for CGRP(8-37).

CGRP receptor antagonists have been tested *in vivo*. For example, as indicated, CGRP(8-37) has been shown to reverse hypotension and tachycardia produced by bacterial lipopolysaccharide (LPS) administration. Therefore, this invention also relates to a therapeutically effective amount of the peptides of the present invention, preferably in a pharmaceutically acceptable buffer such as phosphate buffered solutions including saline as well as other buffered solutions well known in the art of pharmaceutical formulations that can be administered to an animal, including humans, to limit or otherwise inhibit the effects of CGRP binding to one or more CGRP receptors. The peptides can be delivered to the animal using a method that is suitable for the pathology being treated including, but not limited to, intravascular routes of delivery, parenteral routes, where applicable, intramuscular routes, or through the airways using an aerosol, a drip, or the like.

All references and publications cited herein are expressly incorporated by reference into this disclosure. Particular embodiments of this invention will be discussed in detail and reference has been made to possible variations within the scope of this invention. There are a variety of alternative techniques and procedures available to those of skill in the art which would similarly permit one to successfully perform the intended invention.

Example 1 Synthesis of CGRP(8-37) analogues

10

Chemicals and Materials

N- α -Butyloxycarbonyl (Boc) amino acid derivatives were purchased from Bachem (Torrence, CA) and Applied Biosystems (Foster City, CA.). Reactive side chains of amino acids were protected as follows: Arg, mesitylene-2-sulphonyl; Asp, benzyl ester; Cys, *p*-methoxybenzyl; His, benzyloxymethyl; Lys, 2-chlorocarbobenzoxymethyl; Ser, benzyl ether; Thr, benzyl ether; Trp, formyl; Tyr, 2-bromocarbobenzoxymethyl. Different batches of *para*-methylbenzhydrylamine (*p*-MBHA) resin, from Applied Biosystems, were used with substitutions varying from 0.62-0.77 mmol/g. All solvents and reagents for peptide syntheses were peptide synthesis grade from Applied Biosystems and Fisher Biotechnology (Pittsburgh, PA). Thioanisole, ethanedithiol (EDT), *m*-cresol and dimethylsulfide (DMS) were purchased from Aldrich (Milwaukee, WI), trifluoroacetic acid (TFA) and diethyl ether were from Fisher, trifluoromethanesulfonic acid (TFMSA) was from Applied Biosystems. All chemicals were used as supplied. Analytical and semi-preparative reversed-phase high performance liquid chromatography (RP-HPLC) was performed on a Waters Corporation Inc. 625LC instrument. A VYDAC 218TP510 C₁₈ column (1 x 25 cm) supplied by the Nest Group (Southboro, MA.) was used for semi-preparative RP-HPLC. The flow rate was 4 ml/min and the eluant was continuously monitored at 230 nm and collected in 4 ml fractions. Analytical RP-HPLC was performed on a VYDAC 218TP5415 C₁₈, 300Å, column (0.46 x 15 cm) and a

KROMASIL C₈, 100Å column (0.46 x 25 cm). The flow rate was 1 ml/min and the eluant was continuously monitored at 220 nm. Water was obtained from a Barnstead Nanopure system and solvents for HPLC were Optima grade from Fisher. TFA for HPLC was supplied by Pierce (Rockford, IL.). Solvents used for RP-HPLC were,
5 Solvent A: triethylammonium phosphate (100 mM) pH 2.5; Solvent B: is a mixture of acetonitrile in Solvent A (60/40, vol/vol); Solvent C: 0.1% TFA in water; and Solvent D: 0.09% TFA in acetonitrile/water (60/40, vol/vol). Electrospray ionization mass spectrometry (ESI-MS) was performed on all samples at the Nebraska Center for Mass Spectrometry (Lincoln, NE.).

10 2. Solid Phase Peptide Synthesis of all Analogues.

All peptides were made by Merrifield's solid-phase methodology as described previously (Smith, D.D. et al.; *J. Med. Chem.* 36:2536-2541, 1993).

N- α -Boc amino acid derivatives were coupled to *p*-MBHA resin (0.5 meq.) in a four-fold excess using di-isopropylcarbodiimide and hydroxybenzotriazole in N-methylpyrrolidinone. The coupling reactions were monitored by the quantitative
15 ninhydrin test (Sarin, V.K. et al., *Anal. Biochem.* 117: 147-157, 1981).

The Boc protecting group was removed using 33% TFA in dichloromethane (DCM) for one minute and 50% TFA in DCM for 20 minutes. The first 21 amino acid derivatives were single-coupled and the last 15 amino acid
20 derivatives were double-coupled to maintain yields in excess of 99%. After the twentieth coupling, the peptide-resin was dried and half was used for the rest of the synthesis. Once the desired sequences were assembled the N-terminal Boc group was removed as described above and portions of the peptide resin were either benzylated using benzyl bromide (10 eq.) and DIEA (10 eq.) in DCM for six hours
25 or benzoylated using benzoic anhydride (10 eq.) and DIEA (10 eq.) in DCM for one hour. Peptides were freed of their side chain protecting groups and cleaved from the resin by the low-high TFMSA method of Tam (Tam, J.P. et al., *J. Am. Chem. Soc.* 108: 5242-5251, 1986) to yield an ether precipitated, crude product which was purified immediately.

For benzoyl derivatives and analogues, the N-terminal group can be acylated following previously described procedures (Stewart and Young in Solid Phase Peptide Synthesis, 2nd edition p. 73, Pierce Chemical Co. Rockford, Il.) using either the appropriate benzoic anhydride (10 eq.), benzoyl chloride (10 eq.) or sulphonyl chloride (10 eq.) in the presence of DIEA (10eq.) or the appropriate benzoic acid (4 eq.) and coupling reagents such as diisopropyl carbodiimide (4 eq.) and hydroxybenzotriazole (4 eq.). The acylation reaction can be monitored using the known quantitative ninhydrin test. The benzoylated peptides can be cleaved from the resin and purified.

- 10 In one example, 2-chlorobenzoyl-h- α -CGRP(8-37) is synthesized using 0.25 mmol peptide resin treated with TFA/DCM (1/1 vol/vol), washed DCM (5x1min), mixed with DIEA/DCM (5/95 vol/vol) and washed with DCM (5 x 1 min). A solution of 2-chlorobenzoyl chloride (438 mg, 2.5 mmol) and DIEA (0.43 ml, 2.5 mmol) in DCM (20 ml) is mixed with the deprotected peptide-resin for 1
- 15 hour or until the acylation is complete by the quantitative ninhydrin test. The resin is then washed with DCM (5 x 1 min) and dried under reduced pressure. The peptide is cleaved from the resin and purified.

- In another example, benzyl derivatives and analogues can be prepared by alkylating the N-terminal amino group using the appropriate benzyl halide
- 20 (generally bromo or chloro compounds) (10eq.) in the presence of DIEA (10 eq.). These reaction conditions produce a mixture of the mono-benzylated and di-benzylated peptides, that can then be cleaved from the resin and purified.

Purification of Exemplary Analogues

- Analogue 1a: N- α -benzyl-h- α -CGRP (8-37) and analogue dibenzyl-h- α -CGRP(8-*
- 25 *37).* The crude product derived from benzylating 280 mg of peptide resin, was dissolved in solvent A (*supra*) and loaded on to the semi-preparative RP-HPLC column previously equilibrated with a mixture of solvent A and solvent B (76/24 vol/vol). The column was eluted using a linear gradient increasing solvent B composition to 54% over 75 minutes to yield two major products. Fractions

containing the first product to elute from the column were pooled, diluted with an equal volume of water and loaded onto the same semi-preparative RP-HPLC column previously equilibrated with a mixture of solvent C and solvent D (67/33, vol/vol). The product was eluted from the column using a linear gradient increasing solvent D composition to 53% over 50 minutes. Fractions containing the product, as determined by analytical RP-HPLC, were pooled and lyophilized to yield 5.2 mg of a white fluffy powder. The product was >98% pure by analytical RP-HPLC. k' values under isocratic conditions and measured masses, determined by ESI-MS, are listed in Table 1.

Fractions containing the later eluting product were pooled, diluted with an equal volume of water and loaded onto the semi-preparative RP-HPLC column previously equilibrated with a mixture of solvent C and solvent D (64/36, vol/vol). This product was eluted from the column using a linear gradient increasing solvent D composition to 56% over 50 minutes. Fractions containing only the desired product, as determined by analytical RP-HPLC, were pooled and lyophilized to yield 8.5 mg of a fluffy, white powder. The product was > 98% pure by analytical RP-HPLC and identified as dibenzyl-h- α -CGRP(8-37) by ESI-MS. k' values under isocratic conditions and measured masses, determined by ESI-MS, are provided in Table 1.

Analogue 1b: N- α -benzoyl-h- α -CGRP (8-37). The title compound resulting from benzoylation of 200 mg of peptide resin, was purified following the same methods described above for N- α -benzyl-h- α -CGRP (8-37). The product was eluted from the semi-preparative RP-HPLC column using a linear gradient of 31% to 51% solvent B over 50 minutes followed by a linear gradient of 35% to 55% solvent D over 50 minutes, to yield 20 mg of the desired lyophilized product (19%). k' values under isocratic conditions and measured masses, determined by ESI-MS, are listed in Table 1 (below).

Analogue 2a: N- α -benzyl-h- β -CGRP (8-37) and dibenzyl-h- β -CGRP(8-37). N- α -benzyl-h- β -CGRP (8-37) and dibenzyl-h- β -CGRP(8-37) were obtained from benzylating 200 mg of peptide resin and purified following the same methods

described above for N- α -benzyl-h- α -CGRP (8-37). Both products were eluted from the semi-preparative RP-HPLC column using a linear gradient of 27% to 47% solvent B over 50 minutes. The first product to elute from the column was further purified on the semi-preparative column using a linear gradient of 31% to 51% solvent D over 50 minutes to yield 4 mg of a white fluffy powder. The product was > 98% pure by analytical RP-HPLC and identified as N- α -benzyl-h- β -CGRP(8-37) by ESI-MS. k' values under isocratic conditions and measured masses, determined by ESI-MS, are listed in Table 1.

The later eluting product was further purified by semi-preparative RP-HPLC using a linear gradient of 34% to 54% solvent D over 50 minutes to yield 5.3 mg of a white, fluffy powder. The product was > 98% pure by analytical RP-HPLC and identified as dibenzyl-h- β -CGRP(8-37) by ESI-MS. k' values under isocratic conditions and measured masses, determined by ESI-MS are listed in Table 1.

Analogue 2b: N- α -benzoyl-h- β -CGRP (8-37). The title compound, derived from benzoylating 200 mg of peptide resin, was purified following the same methods described above for N- α -benzyl-h- α -CGRP (8-37). The product was eluted from the semi-preparative RP-HPLC column using a linear gradient of 32% to 52% solvent B over 50 minutes followed by a linear gradient of 36% to 56% solvent D over 50 minutes to yield 10 mg of the desired lyophilized product (10%). k' values under isocratic conditions and measured masses, determined by ESI-MS, are listed in Table 1.

Table 1.
Physicochemical Properties of Analogues

	Peptide	ESI-MS		Analytical RP-HPLC (k')	
		Calculated	Observed	^a System 1	^b System 2
5	N- α -benzyl-h- α -CGRP (8-37)	3215.6	3215.6	0.6	5.6 ^c
	N- α -benzoyl-h- α -CGRP (8-37)	3229.6	3229.4	3.3	7.7
	N- α -benzyl-h- β -CGRP (8-37)	3221.0	3221.3	0.7	2.0
10	N- α -benzoyl-h- β -CGRP (8-37)	3233.7	3233.5	4.3	4.1 ^d
	Dibenzyl-h- α -CGRP (8-37)	3305.5	3305.6	1.8	3.8
	Dibenzyl-h- β -CGRP (8-37)	3311.1	3310.7	1.8	4.7

^aVYDAC C₁₈ column (0.46 x 15 cm), 53% solvent C, 47% solvent D, 1 ml./min.

^bKROMASIL C₈ column (0.46 x 25 cm), 52% solvent C, 48% solvent D, 1 ml./min.

15 ^cSame as ^b using 56% solvent C, 44% solvent D. ^dSame as ^b using 50% solvent C, 50% solvent D.

Example 2 CGRP antagonist testing

1. Membrane Preparations

20 Cell membranes were prepared from left circumflex, left anterior descending and right circumflex epicardial coronary arteries dissected from fresh pig
25 hearts obtained from a local slaughterhouse. The arteries were cleaned of surrounding fat and connective tissue. They were cut open exposing the luminal surface and the endothelium was removed by rubbing. Arteries were then cross-cut into thin strips with a razor blade and homogenized in ice-cold 50 mM Tris-HCl buffer, pH 7.4, containing 5
30 mM EDTA (Na₂-Ca salt) using a Polytron (speed setting 5 for 20 sec). The homogenate was centrifuged at 1600 g for 10 min at 4°C. The supernatant was collected, homogenized and crude membranes pelleted by high-speed centrifugation at 50,000 g for 30 min at 4°C. The membrane pellet was reconstituted in ice cold 50 mM Tris-HCl, pH 7.4, containing 100 mM NaCl and 5 mM MgCl₂ and the steps of homogenization and

centrifugation repeated twice as described above. The dried membrane pellet was then stored at -80°C until use. Protein was determined according to the method of Lowry (Lowry, O.H. et al., *J. Biol. Chem.* 193:265-275, 1951)

Radioligand Binding Studies.

- 5 Crude membranes (50µg membrane protein/tube) were incubated with varying concentrations of drugs (h-α-CGRP (8-37), N-benzyl-h-α-CGRP (8-37) or N-benzoyl h-α-CGRP (8-37)) together with 50 pM¹²⁵I-[His¹⁰]-h-α-CGRP for 50 min at 37°C. Incubations were performed in 50 mM Tris-HCl buffer, pH 7.4 containing 5 mM MgCl₂, 100 mM NaCl, 0.1% (w/v) bovine serum albumin and 0.05% (w/v) bacitracin.
- 10 Non-specific binding was defined as binding remaining in the presence of 1 µM h-α-CGRP. Bound ¹²⁵I-h-α-CGRP was separated from free by vacuum filtration (Brandel cell harvester, model MG-48R) through glass fiber filters (Schliecher & Schuell, #32) and counted using a γ counter (LKB Wallac 1277). To reduce non-specific binding of peptides to charged surfaces, glass incubation tubes were coated with Sigmacote and
- 15 glass fiber filters soaked for 60 min in 0.2% (v/v) polyethyleneimine prior to use.

We established several criteria in order for each experiment to be considered as valid data. These criteria stated that specific binding of the radiolabel was greater than or equal to 70% and that the proportion of radiolabel bound to the membrane was less than 10% of the total amount added to the incubation.

20 Relaxation of pig coronary arteries.

- The proximal portion of the left circumflex coronary artery was dissected from pig hearts at a local slaughterhouse and transported in ice-cold Krebs' solution (composition in mM; NaCl 125, KCl 5.5, CaCl₂ 2H₂O 2.5, MgCl₂ 6H₂O 1.2, NaH₂PO₄ 1.25, NaHCO₃ 25, dextrose 11.1, Na₂Ca-EDTA 2H₂O 0.029), equilibrated with 95% O₂/5% CO₂.
- 25 Arteries were cleaned of adhering fat and connective tissue. Rings (2 mm long) were cut and mounted between two stainless-steel pins passed through the lumen of the vessel, then placed in water-jacketed organ baths maintained at 37°C. Rings were bathed in Krebs solution gassed with 95% O₂/5% CO₂, pH 7.4. One pin was connected to a Grass FT.03 force transducer for measurement of isometric tension with a Grass model

7D polygraph (Quincy, MA.). Coronary artery rings were equilibrated at 6g of resting tension (determined to be optimal in previous length-tension experiments by Bockman, C.S. et al., *J. Pharmacol. Exp. Ther.* 267: 1126-1133, 1993) for 30 min and then challenged twice with 45 mM KCl. To measure relaxation, tone was induced in the rings using a submaximal dose of KCl (ca. 15mM) and when the response reached a stable degree of contractile tone, complete cumulative concentration-response relaxation curves for agonists were generated. EC₅₀ values (i.e., the concentration of analog needed to cause one-half of maximal relaxation) were used to quantify the potency of agonists in causing relaxation and were calculated by non-linear regression of all data points on the relaxation concentration-response curve.

Functional determination of antagonist affinity values.

To determine antagonist affinity values, coronary artery rings were prepared, equilibrated and contracted as described above. In some experiments, endogenous CGRP was depleted by incubating rings in Krebs' solution containing 100 μM capsaicin and 10μM indomethacin for three hours. Indomethacin was added to prevent capsaicin-induced contraction of coronary arteries mediated through release of prostaglandins from the adventitia (Franc-Cereceda, A., et al. *Eur. J. Pharmacol.* 142: 235-243, 1987). Rings were then washed extensively for one hour to remove capsaicin and indomethacin. Cumulative concentration-response curves for h-α-CGRP-induced relaxation were generated in all rings, and the rings were then washed and re-equilibrated with Krebs' solution for 60 min. Control rings were incubated with Krebs solution only for 90 min followed by relaxation concentration-response curves for h-α-CGRP. No change in the potency of h-α-CGRP in causing relaxation was observed after the 90 min incubation period in control arteries. Some rings were then incubated with the antagonist, h-α-CGRP (8-37) for 90 min prior to beginning concentration-response curves for h-α-CGRP-induced relaxation. Three adjacent rings from each animal were treated with different concentrations of antagonist. For each concentration of antagonist used, dose-ratios were calculated by dividing the EC₅₀ value for h-α-CGRP-induced relaxation in the

presence of antagonist by its EC_{50} value in the absence of antagonist. Schild plots were constructed and linear regression used to determine the X-intercept (pA_2 value). The slopes of the Schild plots are expressed as the mean $\pm 95\%$ confidence limit. Differences in the slopes of Schild plots were determined by analysis of covariance. The individual pA_2 values were averaged and expressed as mean K_B values by conversion to their antilogs.

The inhibition of binding of the peptides of Table 1, such as ^{125}I -[His¹⁰]-h- α -CGRP (1-37) binding, to membranes prepared from pig coronary arteries is shown in Figure 1A and 1B. Membranes were prepared from the left circumflex, left anterior descending and right circumflex coronary arteries. Competition binding experiments were performed by incubating coronary artery membranes (50 μ g protein/tube) with 50 pM ^{125}I -[His¹⁰]-h- α -CGRP and 16 different concentrations of cold ligand. Nonspecific binding was defined experimentally as the bound radioactivity remaining in the presence of 1 μ M h- α -CGRP or 1 μ M h- β -CGRP. Inhibition is expressed in the figures as percent of ^{125}I -[His¹⁰]-h- α -CGRP binding. The potency of each of these analogs in competing for binding to CGRP receptors was determined from nonlinear regression analysis of all data points on the curve. The rank order of potency for the modified α -CGRP peptides in inhibiting ^{125}I -[His¹⁰]-h- α -CGRP from these binding sites was dibenzyl-h- α -CGRP(8-37) > N-benzoyl-h- α -CGRP(8-37) > N-benzyl-h- α -CGRP(8-37) > H- α -CGRP(8-37). The rank order of potency for the modified β -CGRP peptides in inhibiting ^{125}I -[His¹⁰]-h- α -CGRP from these binding sites was N-benzoyl-h- β -CGRP(8-37) \geq dibenzyl-h- β -CGRP(8-37) > N-benzyl-h- β -CGRP(8-37) > h- β -CGRP(8-37). Each competition curve shown was the mean of 3 or 4 experiments, each using membranes prepared from different animals.

The IC_{50} values for each of h- α -CGRP (8-37) modified peptides and h- β -CGRP (8-37) modified peptides are listed in Table 2 and Table 3, respectively. Since the results are similar for either the α - or β -form, only the α - will be discussed. The modified peptides possessed higher affinity than the CGRP1 receptor selective antagonist, h- α -CGRP (8-37) or h- β -CGRP (8-37). Except for N- α -benzyl-h- α -CGRP (8-37), all of these peptides were able to compete with the radioligand for binding at a single affinity site. In

the case of N- α -benzyl-h- α -CGRP (8-37), This compound appeared to compete for binding to high- and low-affinity binding sites, suggesting the presence of two receptors.

Data were analyzed using non-linear least squares curve fitting (Graphpad Inplot). The concentration of unlabeled peptide required to inhibit the binding of ^{125}I -[His¹⁰]-CGRP from half of these sites (IC_{50}) was taken as the measure of affinity of each of these peptides. Non-specific binding determined using 1 μM h- α -CGRP was not different from non-specific binding determined from treating the minimum value of the competition curve as a fitted parameter. Therefore, in our analysis non-specific binding was defined by the minimum value of the competition curve. Comparisons of one- and two-site fits of binding data were made using an F-test option. In cases where $P < 0.05$, the two-site binding model was accepted as the best fit of the data.

Results of the relaxation studies are illustrated in Figure 2. As provided above, ring segments of pig left circumflex coronary artery were cleaned and mounted in glass chambered organ baths filled with Drebs-Henseleit buffer, pH 7.4 maintained at 37°C. Isometric tension on these ring segments was measured by force transducers and recorded on a polygraph. Cumulative concentration-response curves for h- α CGRP were generated on each segment in the presence and absence of antagonist. The mean concentration response curves from 4 experiments illustrating the rightwards shifts of h- α CGRP-induced relaxation caused by increasing concentrations of h- α CGRP(8-37) (Fig. 2A), N-benzoyl-h- α CGRP(8-37) (Fig. 2B), N-benzyl- α CGRP(8-37) (Fig. 2C) and dibenzyl-h- α CGRP(8-37) (Fig. 2D) are shown. The shifts of the agonist concentration - response curves produced by each concentration of the antagonist were used to calculate dose-ratio values. Linear regression analysis of these dose-ratio values plotted on a log (DR-1) vs. log [antagonist] plot was used to determine the affinity of the antagonist, denoted by the x-intercept of the regression.

N-benzyl-h- α -CGRP (8-37) and N-benzoyl-h- α -CGRP (8-37) both inhibited h- α -CGRP-induced relaxation of isolated pig coronary artery rings. Similar to h- α -CGRP (8-37), increasing concentrations of these two compounds produced increasing rightwards shifts of the h- α -CGRP concentration-response curve. Both compounds had higher

affinity for the CGRP receptor in this tissue. Relative to the K_b of 970 nM for h- α -CGRP (8-37), the K_b for N-benzyl-h- α -CGRP (8-37) was 407 nM and the K_b for N-benzoyl-h- α -CGRP (8-37) was 55 nM. Due to their higher affinity in blocking h- α -CGRP responses than CGRP(8-37), both these antagonists can replace CGRP(8-37).

5

Table 2

Radioligand Binding Data for h- α -CGRP (8-37) modified analogs

	Analogue	Affinity IC₅₀ (nM)	p
	h- α -CGRP (8-37)	14.38±0.38	
10	N-benzyl-h- α -CGRP (8-37)	1.58±0.38	0.004
	N-benzoyl-h- α -CGRP (8-37)	0.27±0.0	<0.0001
	dibenzyl-h- α -CGRP(8-37)	0.22±0.06	<0.0001

15 Data was generated from 4 individual experiments, each using membranes prepared from different animals. p value (binding): is the comparison of mean IC₅₀ value of analog to h- α -CGRP(8-37) or h- β -CGRP(8-37) by students t-test analysis. IC₅₀ refers to the concentration of analog that produces 50% inhibition of specific ¹²⁵I-[His¹⁰]-h- α -CGRP binding from pig coronary artery membranes.

20

Table 2A

Inhibition of Relaxation for h- α -CGRP(8-37) modified analogs

	Peptide Antagonist	K_b nM	p
	h- α -CGRP (8-37)	970.1± 300	
25	N-benzyl-h- α -CGRP (8-37)	118.7± 56.4	<0.0001
	N-benzoyl-h- α -CGRP (8-37)	40.36± 18.95	<0.0001
	dibenzyl-h- α -CGRP(8-37)	29.02± 6.37	<0.0001

30 p value (relaxation) refers to the comparison of mean pA2 value of analog to mean pA2 value of h- α -CGRP(8-37) by analysis of covariance. K_b refers to the affinity of the antagonist determined from its inhibition of h- α -CGRP-induced relaxation of pig coronary artery.

Table 3.
Radioligand Binding Data for h- β -CGRP (8-37) modified analogs

	Analogue	Affinity IC ₅₀ (nM)	P
	h- β -CGRP (8-37)	20.65 \pm 3.96	
5	N-benzyl-h- β -CGRP (8-37)	3.62 \pm 0.8	0.0004
	N-benzoyl-h- β -CGRP (8-37)	0.63 \pm 0.09	<0.0001
	dibenzyl-h- β -CGRP(8-37)	0.73 \pm 0.14	<0.0001

Data was generated from 4 individual experiments, each using membranes prepared from different animals. p value(binding) and IC₅₀ are defined as above, using h- β -CGRP(8-37).

Example 3 Antagonist testing in rats

In vivo testing of these peptides uses anesthetized rats. A cannula is placed in the right carotid artery for measurement of blood pressure and a second cannula is placed in the left femoral vein and is used for injection of peptides into the circulation. CGRP is injected first and a reduction in blood pressure has been reported by others (Fisher et al. *Nature* 305:534-536, 1983). After blood pressure returns to normal a CGRP antagonist is given followed by a second injection of CGRP. Inhibition of the hypotensive effect of CGRP by the antagonist is evidence that the reduction in blood pressure is mediated by CGRP receptors.

While particular embodiments of the invention have been described in detail, it will be apparent to those skilled in the art that these embodiments are exemplary rather than limiting, and the true scope of the invention is that defined in the following claims.